

# IAWA LIST OF MICROSCOPIC FEATURES FOR HARDWOOD IDENTIFICATION

with an Appendix on non-anatomical information

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## PREFACE

This list of microscopic features for hardwood identification is the successor to the 'Standard List of Characters Suitable For Computerized Hardwood Identification' published in 1981 (IAWA Bulletin n.s. 2: 99–145) with an explanation of the coding procedure by R.B. Miller. The 1981 publication greatly stimulated international exchange of information and experience on characters suitable for hardwood identification, and inspired considerable debate on the most desirable coding procedures and identification programs. Therefore, at the IAWA meeting during the XIV International Botanical Congress in Berlin, July 1987, it was decided to revise the 1981 standard list. Because of the continuing developments in computer technology and programming, it was agreed to limit the scope of the new list to definitions, explanatory commentary, and illustrations of wood anatomical descriptors, rather than concentrate on coding procedures.

A new Committee was appointed by the IAWA Council to work towards the new list, and thanks to a substantial grant from the USDA Competitive Research Grants – Wood Utilization Program (Grant No. 88-33541-4081), a workshop was held by the Committee from October 2–7, 1988, in the Department of Wood & Paper Science, North Carolina State University, Raleigh, NC, USA, under the joint auspices of IAWA and IUFRO Division 5. A preliminary list was prepared during the workshop. IAWA members were invited to comment on this list, and these comments helped with the final preparation of the new list. The list presented here was agreed to after review of subsequent drafts and extensive internal consultation between committee members.

Although this list has 163 anatomical and 58 miscellaneous features, it is not a complete list encompassing all the structural patterns that one can encounter in hardwoods. Instead it is intended to be a concise list of features useful for identification purposes. Also, the numbers assigned to each feature in the present list are not meant to be codes for a computer program, but are intended to serve for easy reference, and to help translate data from one program/database to another.

Wood and wood cells are biological elements, formed in trees, shrubs, and climbers to fulfill a physiological or mechanical function. Although there is more discrete diversity in wood structure than in many other plant parts, there is also much continuous variation, and any attempt to classify this diversity into well-defined features has an artificial element. Yet we are confident that in the feature list presented here ambiguity of descriptors has been limited to a minimum, and we hope that all present and future colleagues engaged in wood identification and descriptive wood anatomy will find this list a valuable guide and reference.

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### EXPLANATORY NOTES

**Quantitative Features** — For quantitative features of general applicability (e.g., vessel frequency, tangential vessel lumen diameter, vessel element length, and fibre length), this list includes broad categories for easy use when identifying unknowns, as well as more precise quantitative descriptors (mean, range, standard deviation). When constructing a database the numbers of samples as well as the number of measurements or counts done per sample should be recorded. Different computer programs allow storage of different amounts of information (e.g., all measurements, or just the means, ranges, and standard deviations), and use different algorithms for matching quantitative features. This publication does not recommend a particular program or a particular method for the storage and retrieval of quantitative data, but provides some guidance on how to obtain these data.

**Variable Features and Relative Abundance** — Because of wood's inherent variability, it is inevitable that some features will be well-defined in some samples while absent or ill-defined in other samples of the same species. Accommodating such variability has always been a problem in key construction, and most keys (computerised or otherwise) have provisions for such situations. Describing relative abundance of some features, e.g., prismatic crystals, is also problematic, and textual comments on relative frequency should be added to a description or database. In this list of descriptors, some features apply only when the characteristic is of common occurrence. For such features, the illustrations and examples are intended to help interpret 'common'. Although many keys have used these same features accompanied by the same qualifier 'common', there have been no extensive systematic analyses to determine what per cent occurrence constitutes 'common'. Therefore no quantitative criteria for 'common' have been offered in this list.

## LIST OF FEATURES

## Name

## ANATOMICAL FEATURES

**Growth rings** — p. 234

1. Growth ring boundaries distinct
2. Growth ring boundaries indistinct or absent

**Vessels** — p. 236*Porosity* — p. 236

3. Wood ring-porous
4. Wood semi-ring-porous
5. Wood diffuse-porous

*Vessel arrangement* — p. 238

6. Vessels in tangential bands
7. Vessels in diagonal and/or radial pattern
8. Vessels in dendritic pattern

*Vessel groupings* — p. 242

9. Vessels exclusively solitary (90% or more)
10. Vessels in radial multiples of 4 or more common
11. Vessel clusters common

*Solitary vessel outline* — p. 244

12. Solitary vessel outline angular

*Perforation plates* — p. 246

13. Simple perforation plates
14. Scalariform perforation plates
  15. Scalariform perforation plates with  $\leq 10$  bars
  16. Scalariform perforation plates with 10–20 bars
  17. Scalariform perforation plates with 20–40 bars
  18. Scalariform perforation plates with  $\geq 40$  bars
19. Reticulate, foraminated, and/or other types of multiple perforation plates

*Intervessel pits: arrangement and size* — p. 250

20. Intervessel pits scalariform
21. Intervessel pits opposite
22. Intervessel pits alternate
23. Shape of alternate pits polygonal
24. Minute —  $\leq 4 \mu\text{m}$
25. Small — 4–7  $\mu\text{m}$
26. Medium — 7–10  $\mu\text{m}$
27. Large —  $\geq 10 \mu\text{m}$
28. Range of intervessel pit size ( $\mu\text{m}$ )

*Vestured pits* — p. 252

29. Vestured pits



*Vessel-ray pitting* — p. 253

30. Vessel-ray pits with distinct borders; similar to intervessel pits in size and shape throughout the ray cell
31. Vessel-ray pits with much reduced borders to apparently simple: pits rounded or angular
32. Vessel-ray pits with much reduced borders to apparently simple: pits horizontal (scaleriform, gash-like) to vertical (palisade)
33. Vessel-ray pits of two distinct sizes or types in the same ray cell
34. Vessel-ray pits unilaterally compound and coarse (over 10  $\mu\text{m}$ )
35. Vessel-ray pits restricted to marginal rows

*Helical thickenings* — p. 256

36. Helical thickenings in vessel elements present
  37. Helical thickenings throughout body of vessel element
  38. Helical thickenings only in vessel element tails
  39. Helical thickenings only in narrower vessel elements

*Tangential diameter of vessel lumina* — p. 258

Mean tangential diameter of vessel lumina

40.  $\leq 50 \mu\text{m}$
41. 50–100  $\mu\text{m}$
42. 100–200  $\mu\text{m}$
43.  $\geq 200 \mu\text{m}$
44. Mean, +/- Standard Deviation, Range,  $n = x$
45. Vessels of two distinct diameter classes, wood not ring-porous

*Vessels per square millimetre* — p. 259

46.  $\leq 5$  vessels per square millimetre
47. 5–20 vessels per square millimetre
48. 20–40 vessels per square millimetre
49. 40–100 vessels per square millimetre
50.  $\geq 100$  vessels per square millimetre
51. Mean, +/- Standard Deviation, Range,  $n = x$

*Mean vessel element length* — p. 259

52.  $\leq 350 \mu\text{m}$
53. 350–800  $\mu\text{m}$
54.  $\geq 800 \mu\text{m}$
55. Mean, +/- Standard Deviation, Range,  $n = x$

*Tyloses and deposits in vessels* — p. 259

56. Tyloses common
57. Tyloses sclerotic
58. Gums and other deposits in heartwood vessels

*Wood vesselless* — p. 262

59. Wood vesselless

**Tracheids and fibres** — p. 262

60. Vascular/vasicentric tracheids present

*Ground tissue fibres* — p. 264

61. Fibres with simple to minutely bordered pits
62. Fibres with distinctly bordered pits
63. Fibre pits common in both radial and tangential walls
64. Helical thickenings in ground tissue fibres

*Septate fibres and parenchyma-like fibre bands* — p. 266

65. Septate fibres present
66. Non-septate fibres present
67. Parenchyma-like fibre bands alternating with ordinary fibres

*Fibre wall thickness* — p. 268

68. Fibres very thin-walled
69. Fibres thin- to thick-walled
70. Fibres very thick-walled

*Mean fibre lengths* — p. 269

71.  $\leq 900 \mu\text{m}$
72. 900–1600  $\mu\text{m}$
73.  $\geq 1600 \mu\text{m}$
74. Mean, +/- Standard Deviation, Range,  $n = x$

**Axial parenchyma** — p. 270

75. Axial parenchyma absent or extremely rare

*Apotracheal axial parenchyma* — p. 270

76. Axial parenchyma diffuse
77. Axial parenchyma diffuse-in-aggregates

*Paratracheal axial parenchyma* — p. 272

78. Axial parenchyma scanty paratracheal
79. Axial parenchyma vasicentric
80. Axial parenchyma aliform
  81. Axial parenchyma lozenge-aliform
  82. Axial parenchyma winged-aliform
83. Axial parenchyma confluent
84. Axial parenchyma unilateral paratracheal

*Banded parenchyma* — p. 276

85. Axial parenchyma bands more than three cells wide
86. Axial parenchyma in narrow bands or lines up to three cells wide
87. Axial parenchyma reticulate
88. Axial parenchyma scalariform
89. Axial parenchyma in marginal or in seemingly marginal bands

*Axial parenchyma cell type/strand length* — p. 280

90. Fusiform parenchyma cells
  91. Two cells per parenchyma strand
  92. Four (3–4) cells per parenchyma strand
  93. Eight (5–8) cells per parenchyma strand
  94. Over eight cells per parenchyma strand
95. Unlignified parenchyma

**Rays** — p. 282*Ray width* — p. 282

- 96. Rays exclusively uniseriate
- 97. Ray width 1 to 3 cells
- 98. Larger rays commonly 4- to 10-seriate
- 99. Larger rays commonly > 10-seriate
- 100. Rays with multiseriate portion(s) as wide as uniseriate portions

*Aggregate rays* — p. 284

- 101. Aggregate rays

*Ray height* — p. 284

- 102. Ray height > 1 mm

*Rays of two distinct sizes* — p. 286

- 103. Rays of two distinct sizes

*Rays: cellular composition* — p. 288

- 104. All ray cells procumbent
- 105. All ray cells upright and/or square
- 106. Body ray cells procumbent with one row of upright and/or square marginal cells
- 107. Body ray cells procumbent with mostly 2–4 rows of upright and/or square marginal cells
- 108. Body ray cells procumbent with over 4 rows of upright and/or square marginal cells
- 109. Rays with procumbent, square and upright cells mixed throughout the ray

*Sheath cells* — p. 292

- 110. Sheath cells

*Tile cells* — p. 292

- 111. Tile cells

*Perforated ray cells* — p. 294

- 112. Perforated ray cells

*Disjunctive ray parenchyma cell walls* — p. 294

- 113. Disjunctive ray parenchyma cell walls

*Rays per millimetre* — p. 296

- 114.  $\leq 4$  / mm
- 115. 4–12 / mm
- 116.  $\geq 12$  / mm

*Wood rayless* — p. 297

- 117. Wood rayless

**Storied structure** — p. 298

- 118. All rays storied
- 119. Low rays storied, high rays non-storied.
- 120. Axial parenchyma and/or vessel elements storied
- 121. Fibres storied
- 122. Rays and/or axial elements irregularly storied
- 123. Number of ray tiers per axial mm

**Secretory elements and cambial variants** — p. 300*Oil and mucilage cells* — p. 300

- 124. Oil and/or mucilage cells associated with ray parenchyma
- 125. Oil and/or mucilage cells associated with axial parenchyma
- 126. Oil and/or mucilage cells present among fibres

*Intercellular canals* — p. 302

- 127. Axial canals in long tangential lines
- 128. Axial canals in short tangential lines
- 129. Axial canals diffuse
- 130. Radial canals
- 131. Intercellular canals of traumatic origin

*Tubes / tubules* — p. 306

- 132. Laticifers or tanniferous tubes

*Cambial variants* — p. 308

- 133. Included phloem, concentric
- 134. Included phloem, diffuse
- 135. Other cambial variants

**Mineral inclusions** — p. 310*Prismatic crystals* — p. 310

- 136. Prismatic crystals present
  - 137. Prismatic crystals in upright and/or square ray cells
  - 138. Prismatic crystals in procumbent ray cells
  - 139. Prismatic crystals in radial alignment in procumbent ray cells
  - 140. Prismatic crystals in chambered upright and/or square ray cells
  - 141. Prismatic crystals in non-chambered axial parenchyma cells
  - 142. Prismatic crystals in chambered axial parenchyma cells
  - 143. Prismatic crystals in fibres

*Druses* — p. 313

- 144. Druses present
  - 145. Druses in ray parenchyma cells
  - 146. Druses in axial parenchyma cells
  - 147. Druses in fibres
  - 148. Druses in chambered cells

*Other crystal types* — p. 313

- 149. Raphides
- 150. Acicular crystals
- 151. Styloids and/or elongate crystals
- 152. Crystals of other shapes (mostly small)
- 153. Crystal sand

*Other diagnostic crystal features* — p. 315

- 154. More than one crystal of about the same size per cell or chamber
- 155. Two distinct sizes of crystals per cell or chamber
- 156. Crystals in enlarged cells
- 157. Crystals in tyloses
- 158. Cystoliths

*Silica* — p. 318

159. Silica bodies present
  160. Silica bodies in ray cells
  161. Silica bodies in axial parenchyma cells
  162. Silica bodies in fibres
163. Vitreous silica

**APPENDIX — Non-anatomical information — p. 321***Geographical distribution* — p. 321

164. Europe and temperate Asia (Brazier and Franklin region 74)
  165. Europe, excluding Mediterranean
  166. Mediterranean including Northern Africa and Middle East
  167. Temperate Asia (China), Japan, USSR
168. Central South Asia (Brazier and Franklin region 75)
  169. India, Pakistan, Sri Lanka
  170. Burma
171. Southeast Asia and the Pacific (Brazier and Franklin region 76)
  172. Thailand, Laos, Vietnam, Cambodia (Indochina)
  173. Indomalaysia: Indonesia, Philippines, Malaysia, Brunei, Papua New Guinea, and Solomon Islands
  174. Pacific Islands (including New Caledonia, Samoa, Hawaii, and Fiji)
175. Australia and New Zealand (Brazier and Franklin region 77)
  176. Australia
  177. New Zealand
178. Tropical mainland Africa and adjacent islands (Brazier and Franklin region 78)
  179. Tropical Africa
  180. Madagascar & Mauritius, Réunion & Comores
181. Southern Africa (south of the Tropic of Capricorn) (Brazier and Franklin region 79)
182. North America, north of Mexico (Brazier and Franklin region 80)
183. Neotropics and temperate Brazil (Brazier and Franklin region 81)
  184. Mexico and Central America
  185. Caribbean
  186. Tropical South America
  187. Southern Brazil
188. Temperate South America including Argentina, Chile, Uruguay, and S. Paraguay (Brazier and Franklin region 82)

*Habit* — p. 321

189. Tree
190. Shrub
191. Vine/liana

*Wood of commercial importance* — p. 322

192. Wood of commercial importance

*Specific gravity* — p. 322

193. Basic specific gravity low,  $\leq 0.40$
194. Basic specific gravity medium,  $0.40-0.75$
195. Basic specific gravity high,  $\geq 0.75$

*Heartwood colour* — p. 323

- 196. Heartwood colour darker than sapwood colour
- 197. Heartwood basically brown or shades of brown
- 198. Heartwood basically red or shades of red
- 199. Heartwood basically yellow or shades of yellow
- 200. Heartwood basically white to grey
- 201. Heartwood with streaks
- 202. Heartwood not as above

*Odour* — p. 325

- 203. Distinct odour

*Heartwood fluorescence* — p. 325

- 204. Heartwood fluorescent

*Water & ethanol extracts: fluorescence & colour* — p. 326

- 205. Water extract fluorescent
- 206. Water extract basically colourless to brown or shades of brown
- 207. Water extract basically red or shades of red
- 208. Water extract basically yellow or shades of yellow
- 209. Water extract not as above
- 210. Ethanol extract fluorescent
- 211. Ethanol extract basically colourless to brown or shades of brown
- 212. Ethanol extract basically red or shades of red
- 213. Ethanol extract basically yellow or shades of yellow
- 214. Ethanol extract not as above

*Froth test* — p. 327

- 215. Froth test positive

*Chrome Azurol-S test* — p. 328

- 216. Chrome Azurol-S test positive

*Burning splinter test* — p. 328

- 217. Splinter burns to charcoal
- 218. Splinter burns to a full ash: Colour of ash bright white
- 219. Splinter burns to a full ash: Colour of ash yellow-brown
- 220. Splinter burns to a full ash: Colour of ash other than above
- 221. Splinter burns to a partial ash